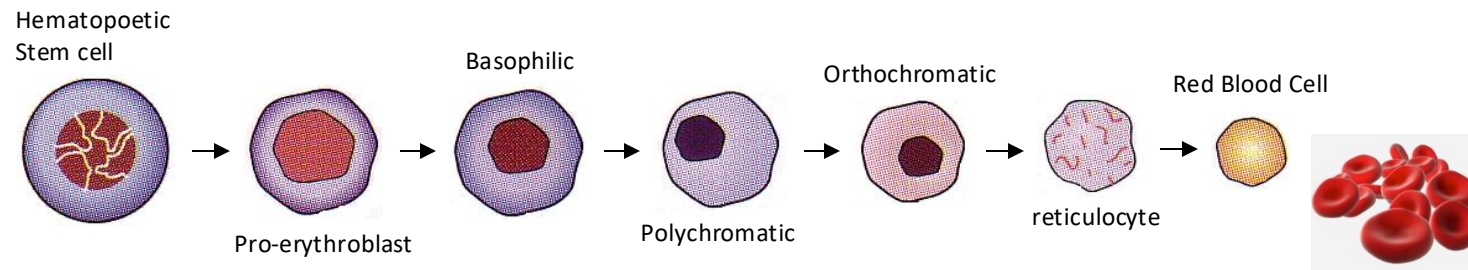
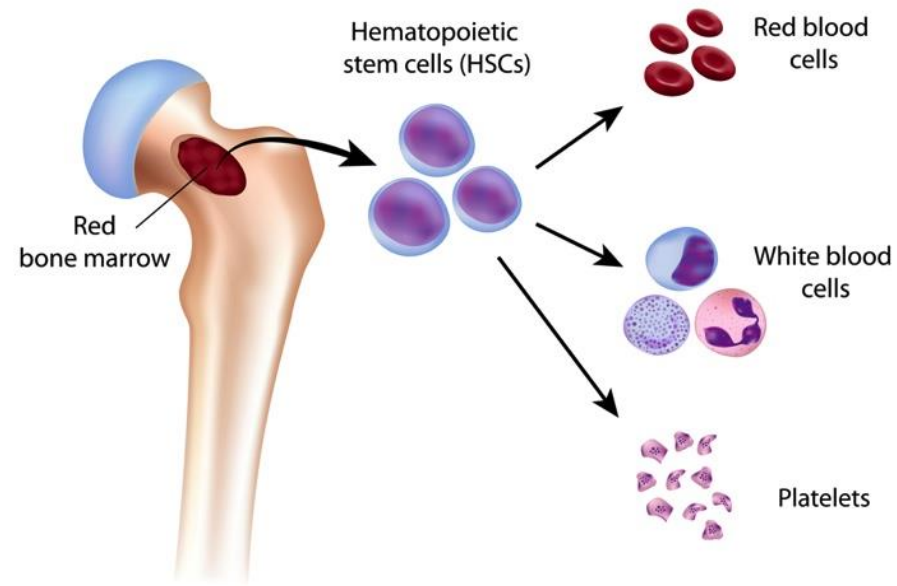


# Genetic Engineering of Blood Products (RBCs)

Jan Frayne & Ash Toyne  
University of Bristol, UK

- Growing RBCs in lab
- Sustainable erythroid lines as source of RBCs
- Genetic engineering of RBCs
- RBCs as drug delivery and targeting agents

# Erythropoiesis in vivo



# Primary aim of in vitro grown RBCs for unmet clinical need

MDS  
Transfusion-dependent patients  
Rare blood types  
Younger cells – reduce transfusion frequency and dose, reduced infectious risks  
Research tool to study erythropoiesis in health and disease

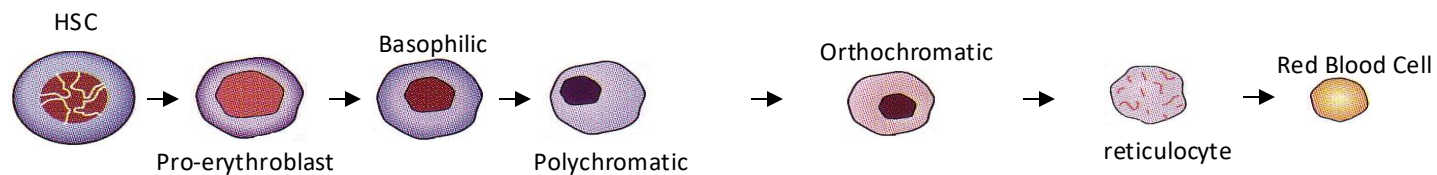
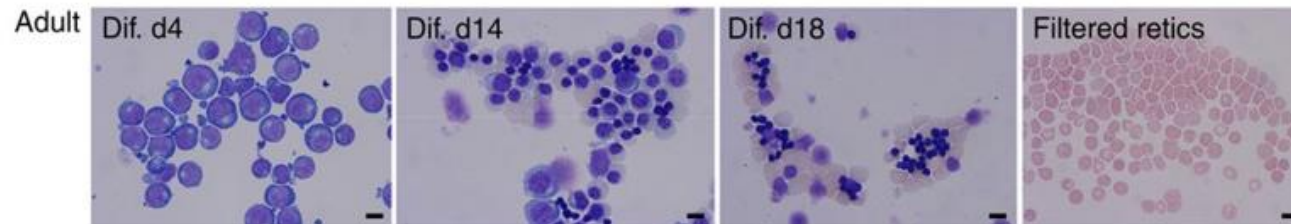
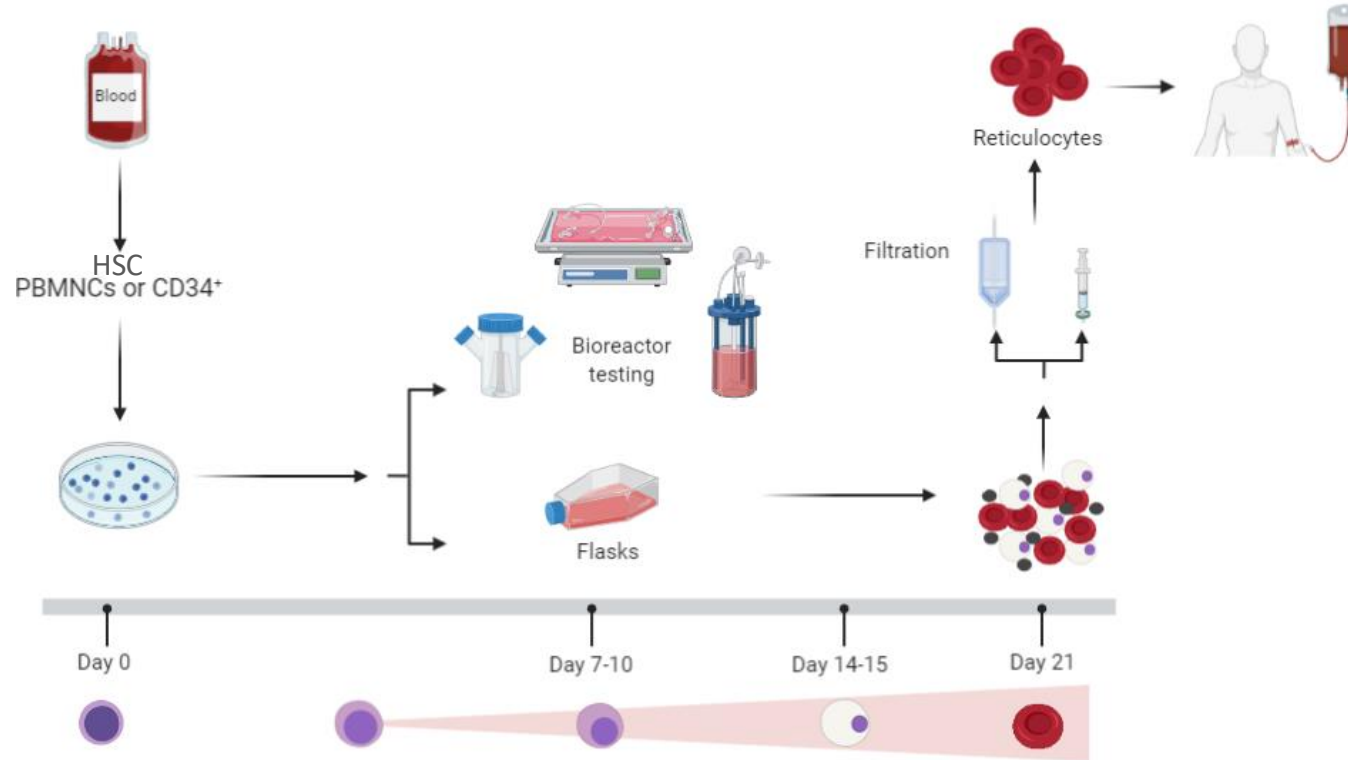
} erythrocyte alloimmunization, iron overload

## Progenitors for *in vitro* generation of Red Blood Cells

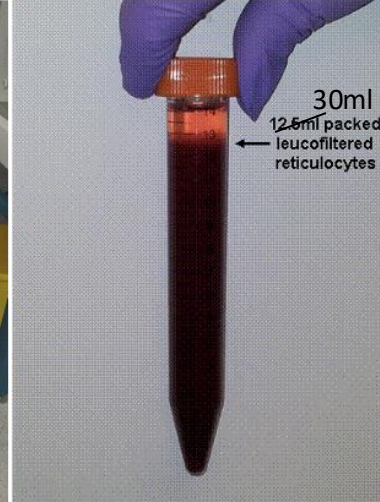


- Bone marrow HSC
- Peripheral blood HSC
- Cord blood HSC
- iPSCs
- ESCs

# *In vitro* culture of red blood cells



# Growing cells at scale under GMP



Good Manufacturing Practice (GMP) Compliant



Images courtesy of Sabine Taylor, Nicky Cogan and Prof. David Anstee (NHSBT)

**RESTORE: Recovery and Survival of Stem Cell Originated Red Cells**

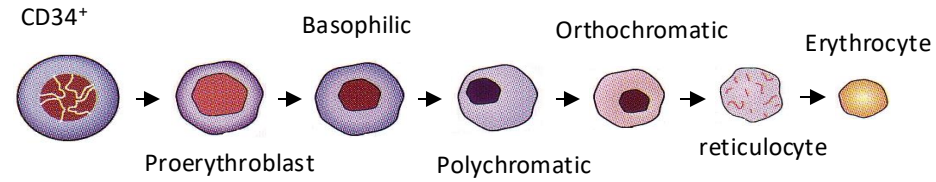
- Phase 1
- Allogeneic



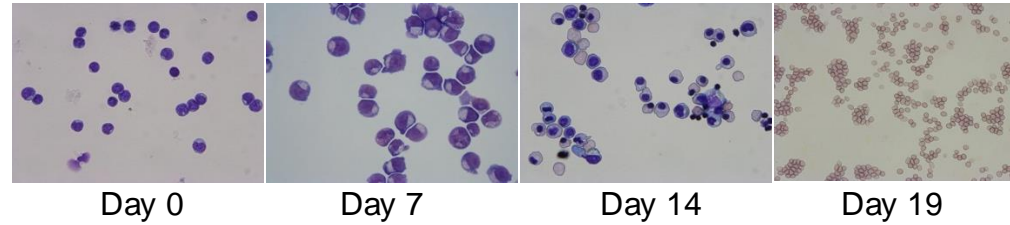
Led by Prof Cedric Ghevaert and Dr Rebecca Cardigan



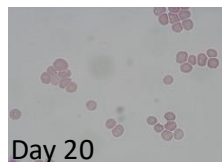
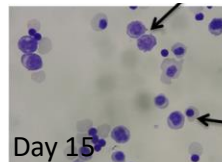
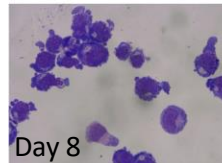
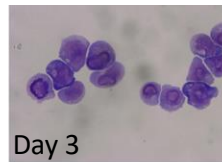
# Hurdles with cultures



Adult PB CD34<sup>+</sup> cells



Cord Blood CD34<sup>+</sup>

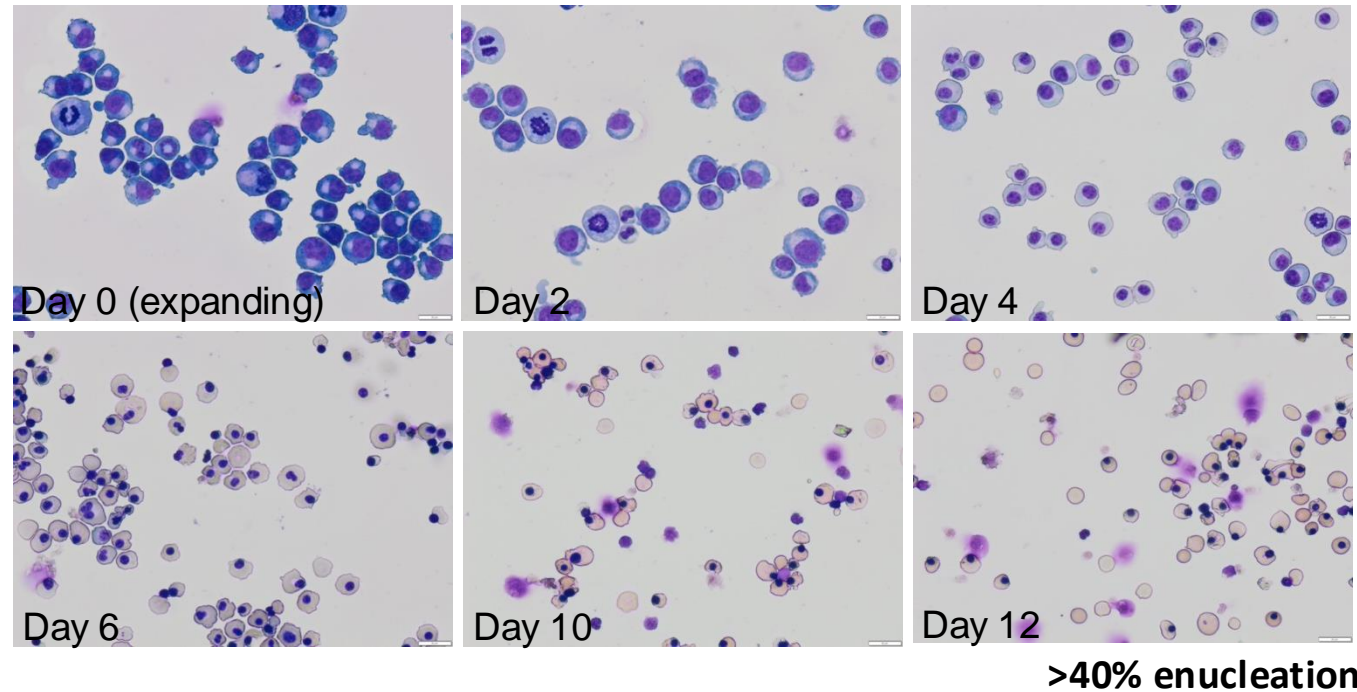
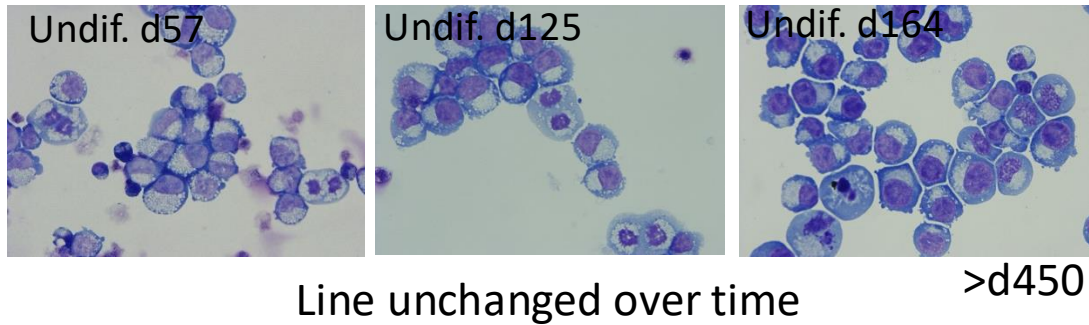
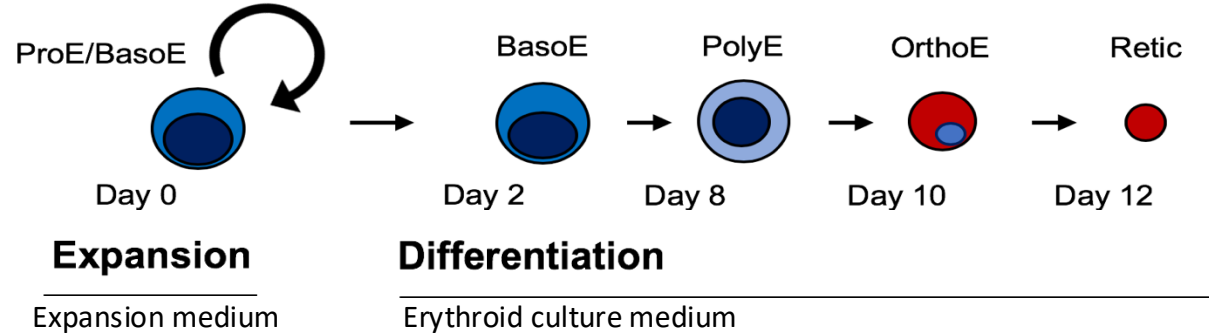
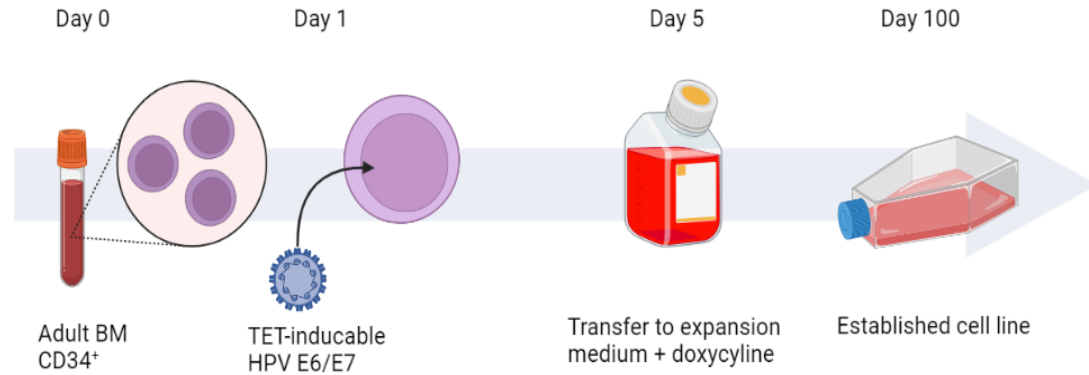


Scalability

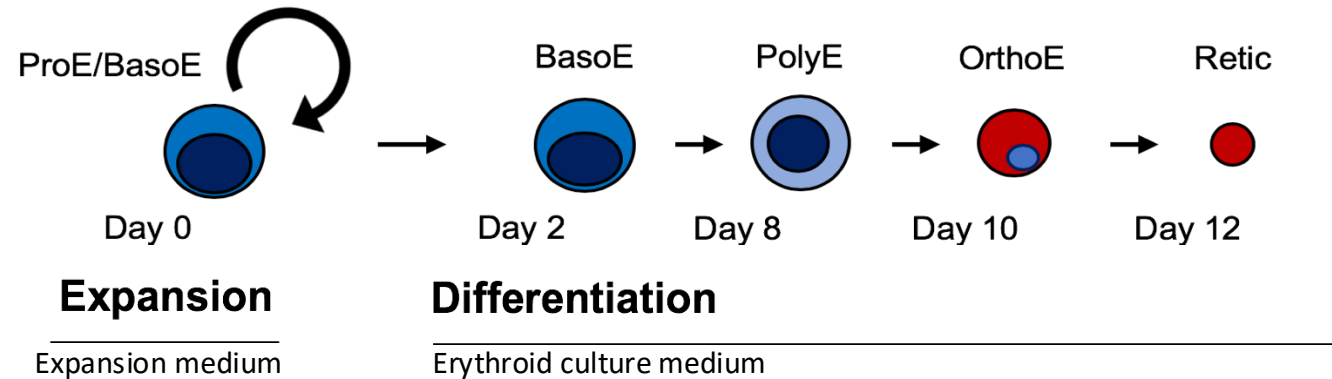
Repeat HSC donations

Available blood groups

# Creating immortalised adult erythroid line (BEL-A)



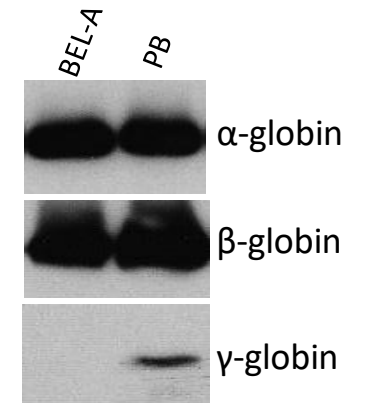
# Differentiated BEL-A cells have adult phenotype



## Extensively Characterised

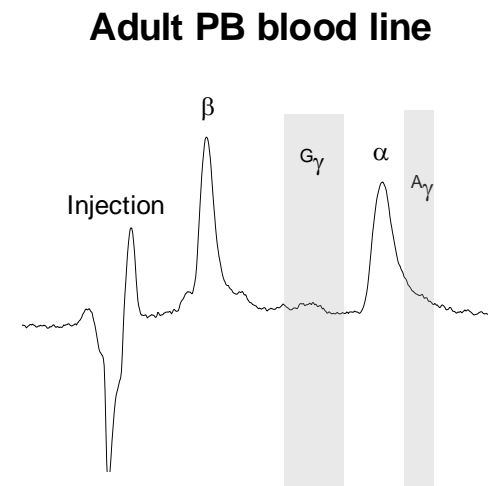
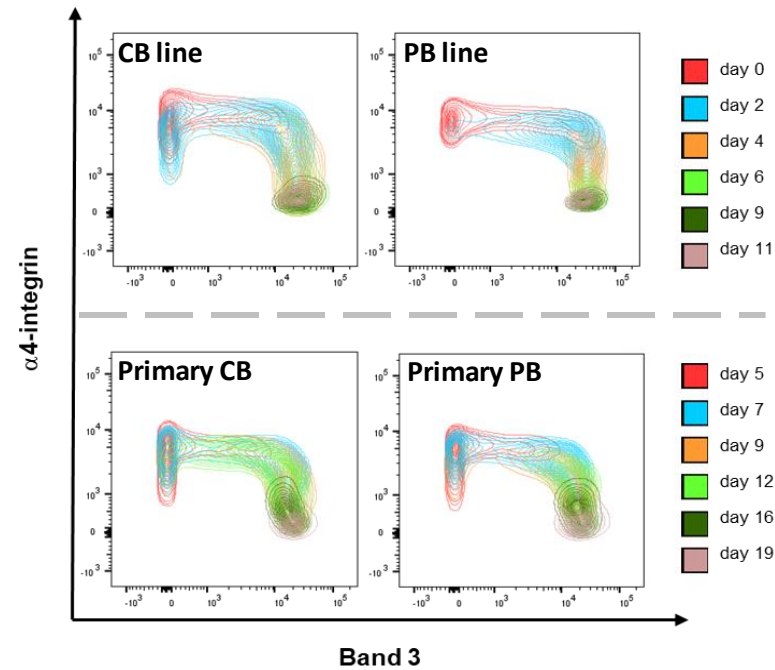
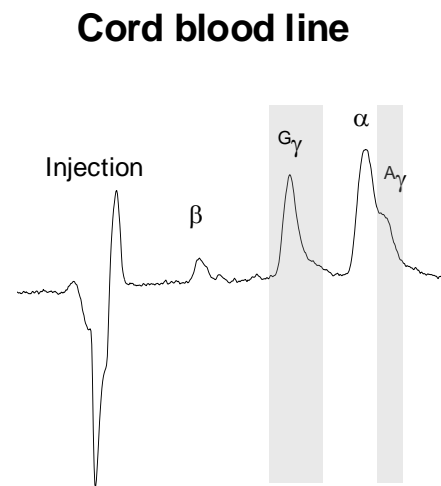
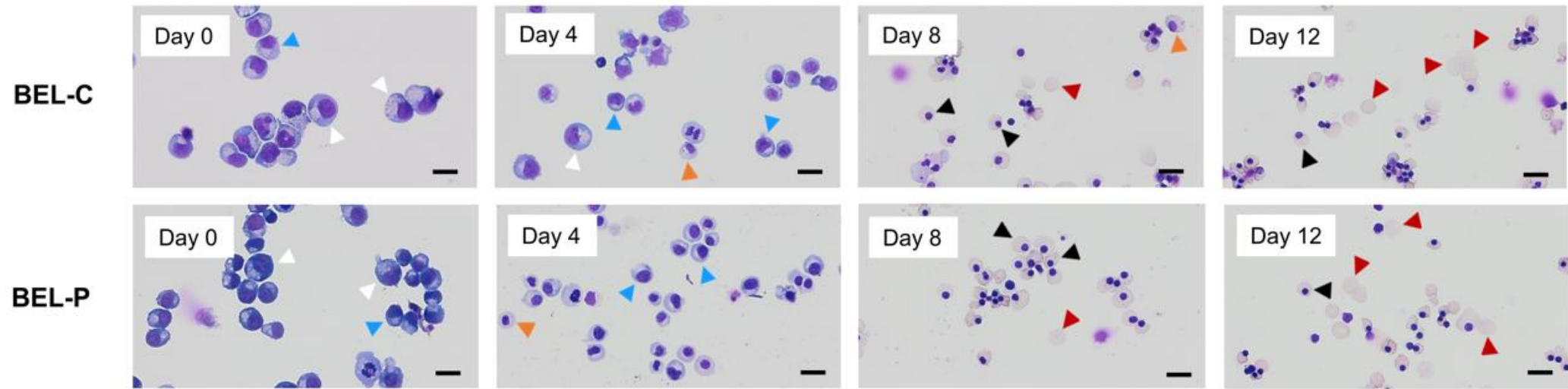
- Trakarnsanga *et al* Nat Comm 2017
- Daniels *et al* Haematologica 2019

- First immortalised adult human erythroid line, recapitulates adult erythropoiesis, produces reticulocytes
- Do not require repeat collection of donor stem cells
- 12 different immortalised erythroid cell lines using same methodology – from BM, PB, CB CD34<sup>+</sup> cells
- Create with chosen blood group phenotype – line has phenotype of the original donor
- **Provides a scalable, sustainable source of red blood cells**

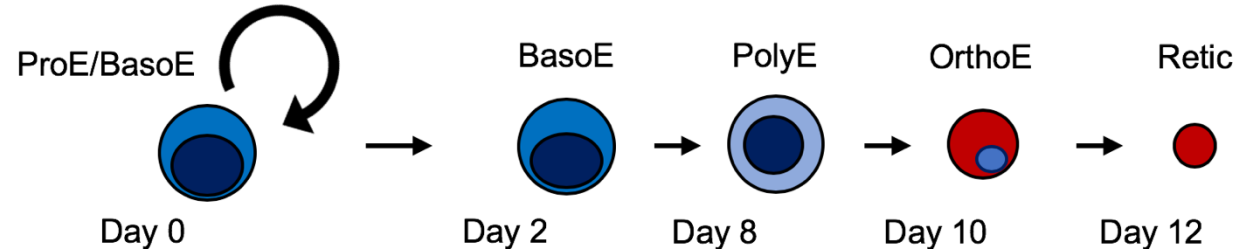




# Lines from more accessible Peripheral & Cord Blood CD34+ recapitulate phenotype of donor erythroid cells



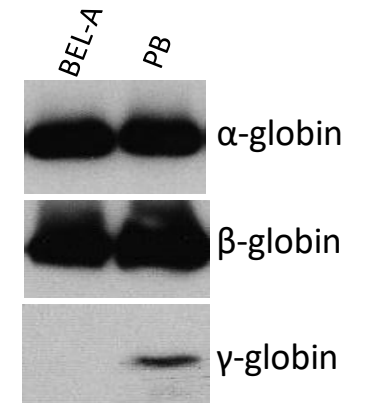
# Differentiated BEL-A cells have adult phenotype



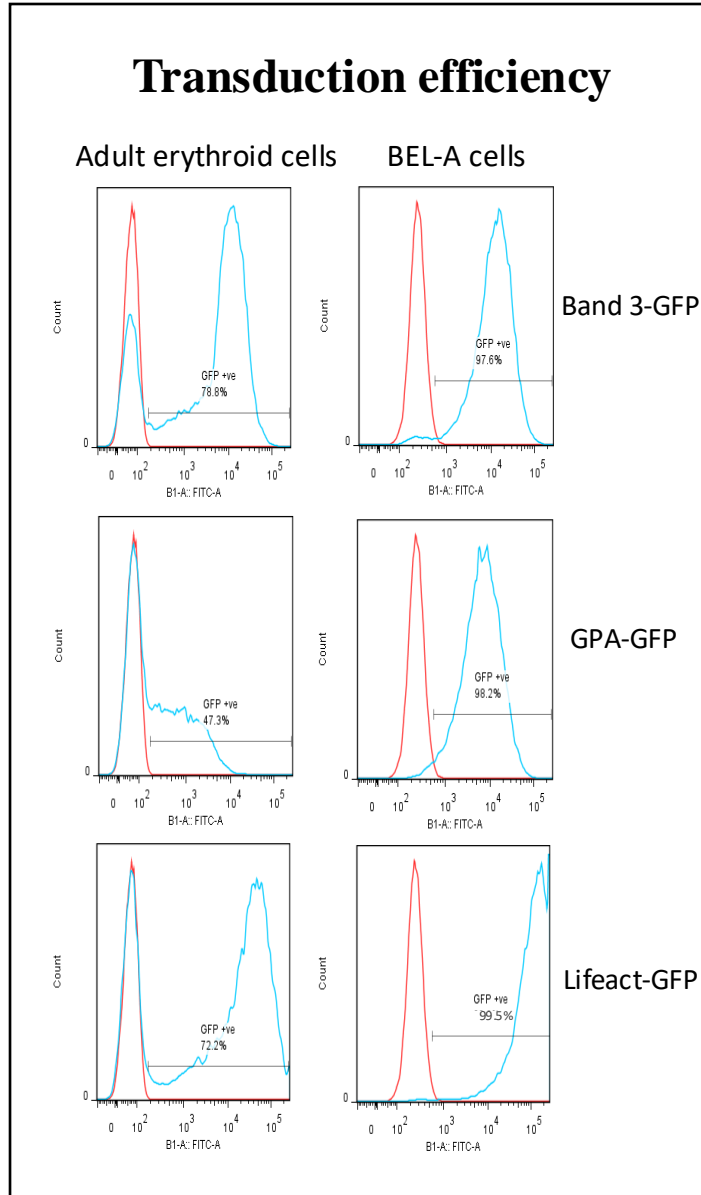
## Extensively Characterised

- Trakarnsanga *et al* Nat Comm 2017
- Daniels *et al* Haematologica 2019

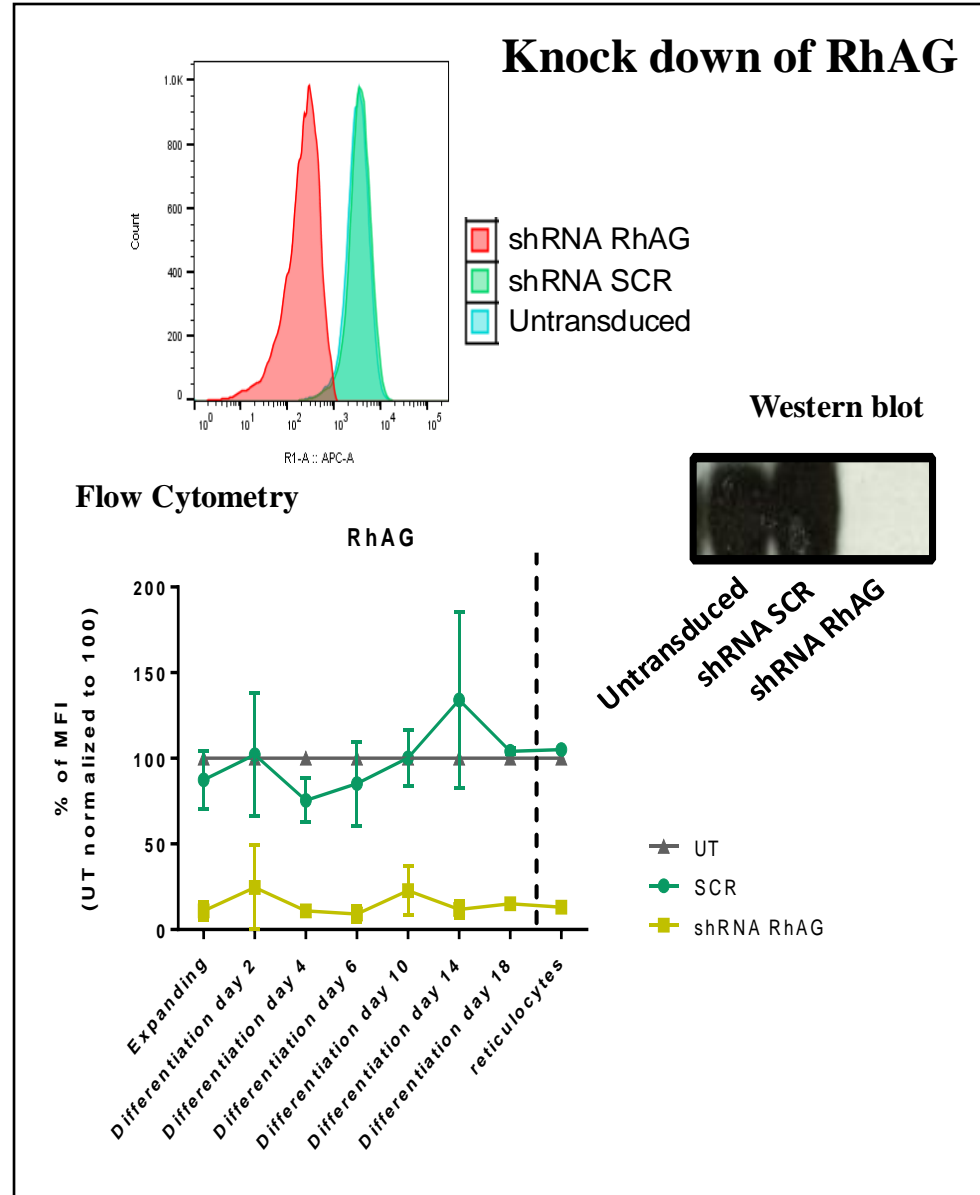
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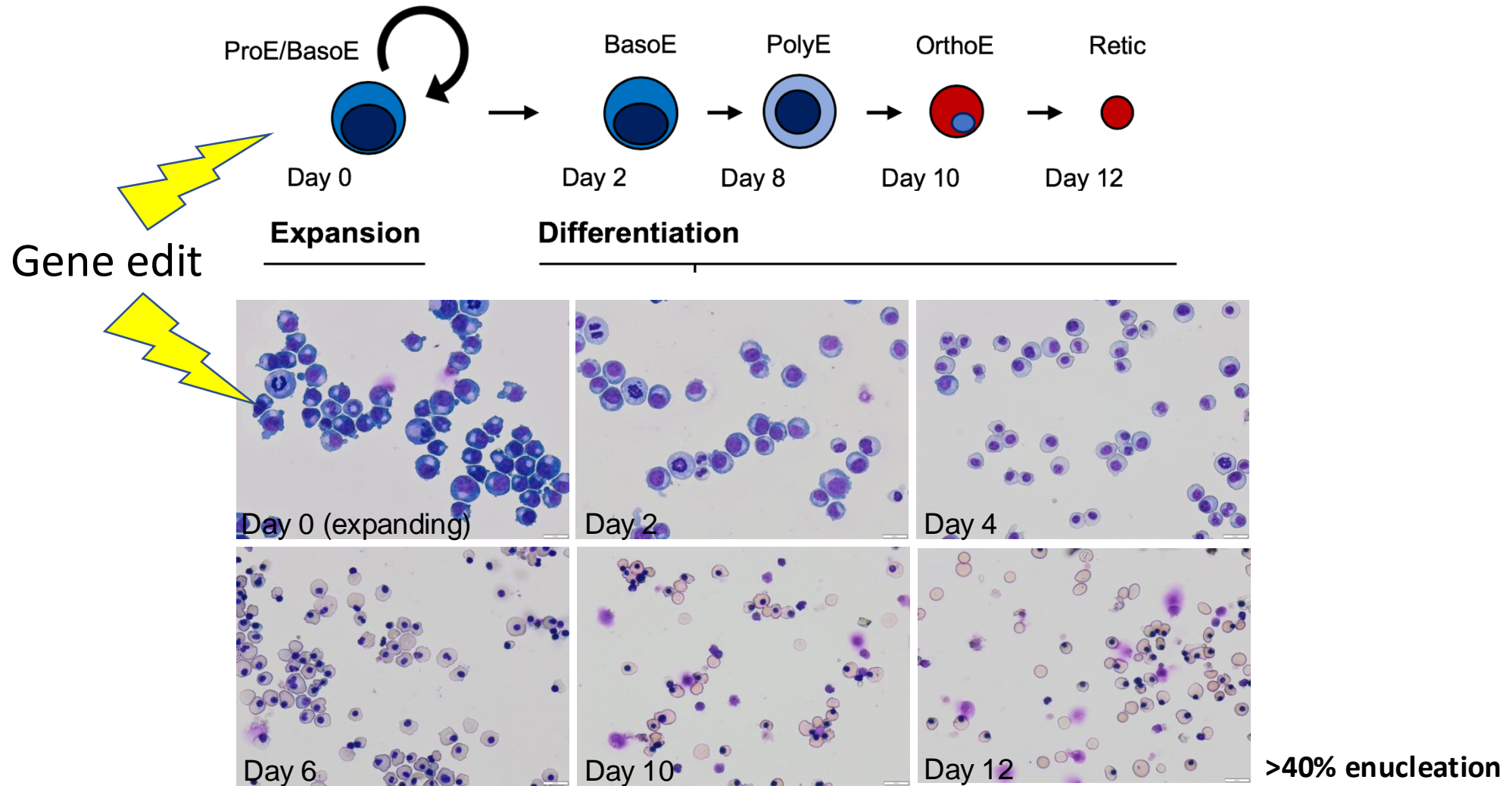
# Overexpression



# shRNA knock down



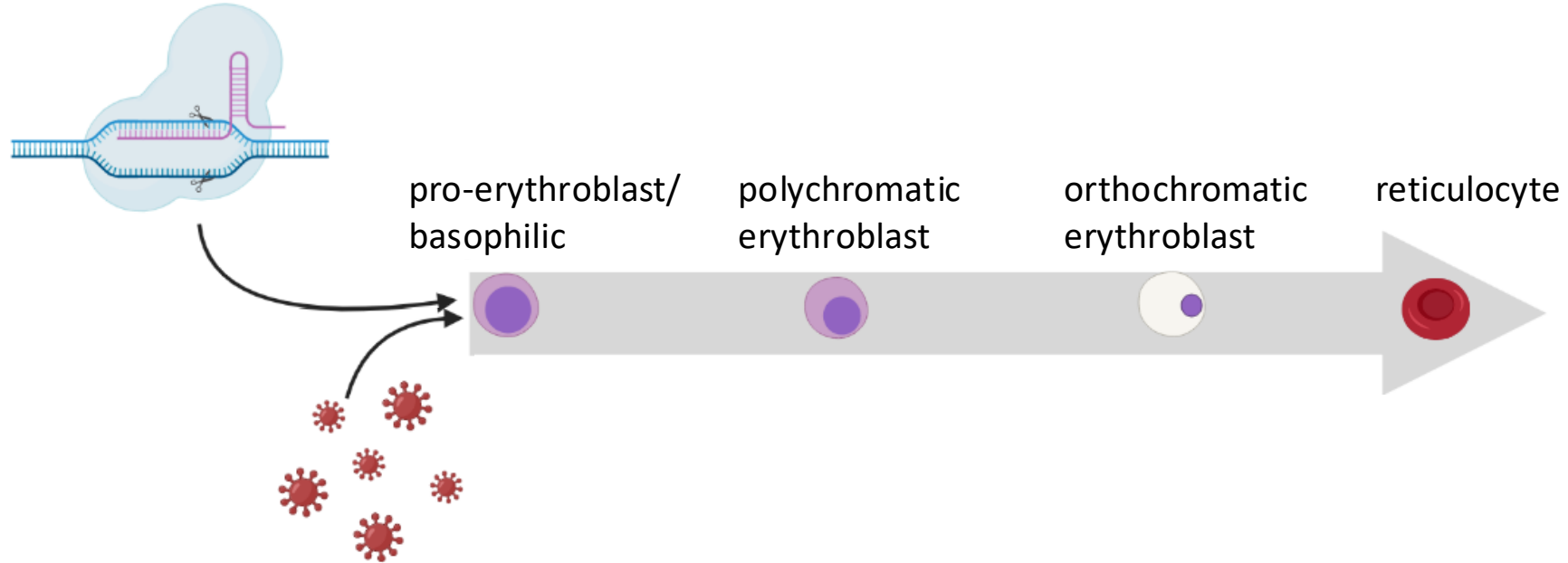
# Platform for CRISPR gene editing of BEL-A cells



# Platform for CRISPR gene editing of BEL-A cells

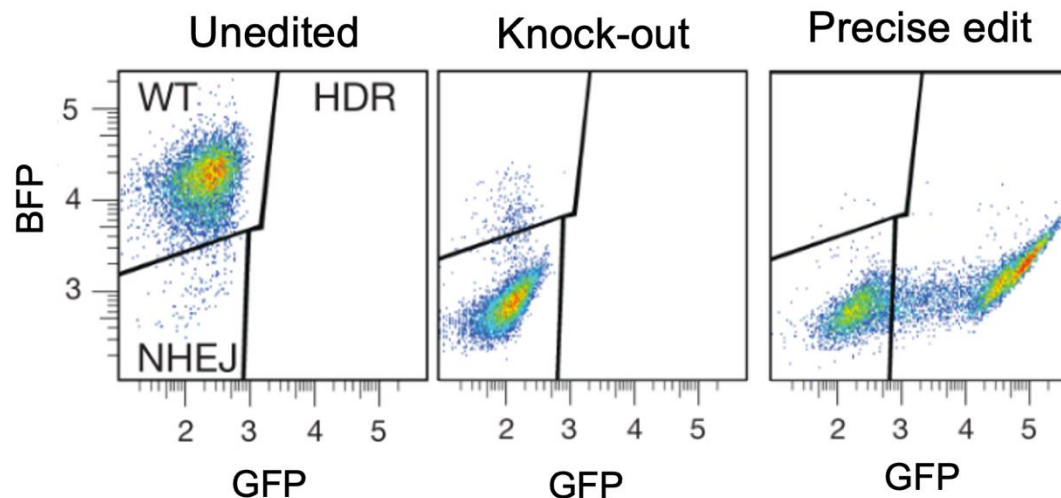
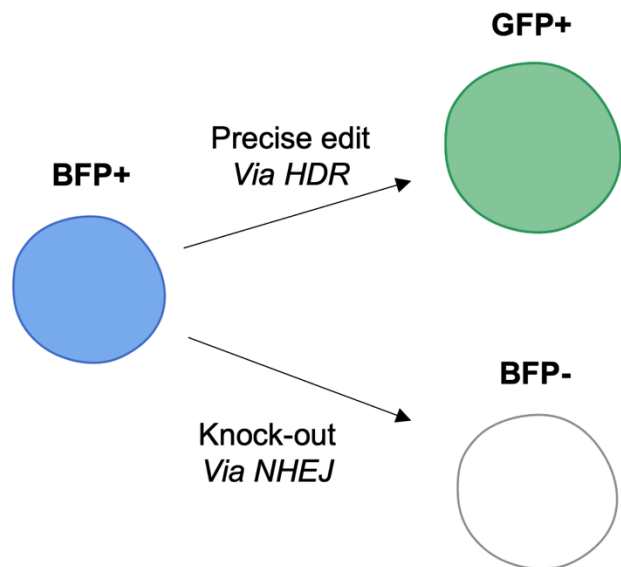
CRISPR gene KO and introduction of specific mutations and tags (and CRISPRa)

Lenti → nucleofection of plasmids → nucleofection of RNPs



Lentiviral over expression and shRNA knock down

# CRISPR Fluorescent Conversion Assay to validate reagents to improve HDR efficiency



Up to 70%

Up to 50% endogenous genes

BFP: ...CCACCCTGACCCATGGCGTGCAGTGCTTCAGCCGCTA...  
 GFP: ...CCACCCTGACGTACGGCGTGCAGTGCTTCAGCCGCTA...

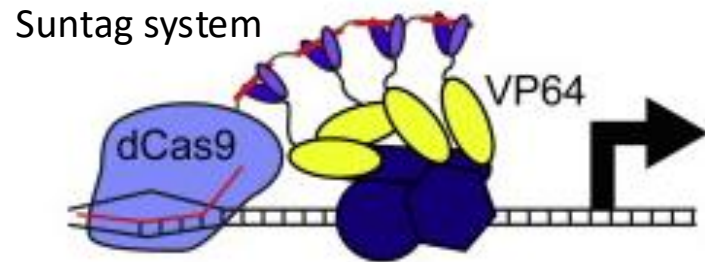
3 bp edit from BFP to GFP

# Applications of BEL-A gene editing

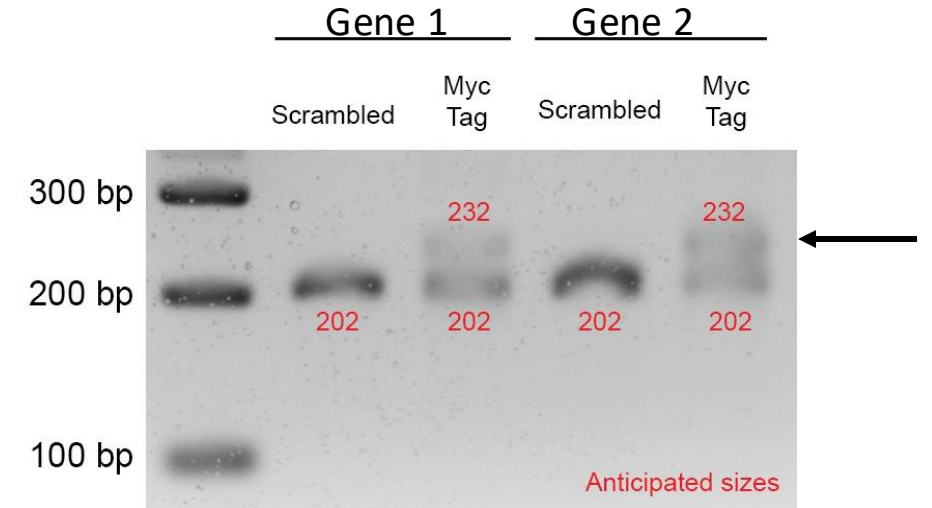
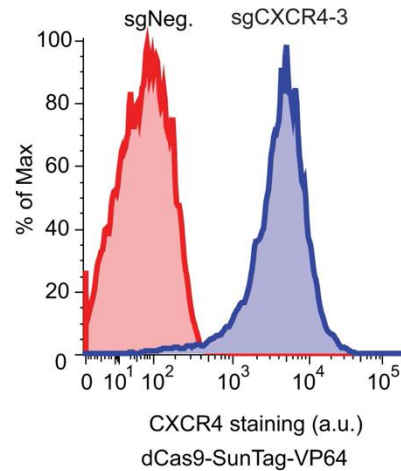
## RBC disease lines from BEL-A as research tools and drug screening platforms

- $\beta$ -thalassemia lines CD41/42 –TTCT, IVS1-1, IVS1-100
- $\alpha$ -thalassemia major lines
- Sickle cell disease lines  $\beta$ 6Glu->Val
- CDA IV lines E325K KLF1

## CRISPRa to induce/increase expression of endogenous genes in BEL-A



Tanenbaum et al Cell 159:635-646



myc tag introduced at 5' of genes

# Improving compatibility and properties of BEL-A erythroid cells

Improved compatibility or  
make more “Universal blood”

Improved efficiency



Improved survival/storage characteristics

Improved oxygen carrying capacity and delivery



# Defining the clinical requirement

Survey for problematic transfusion requirements where matched blood could not be fulfilled or supplies scarce

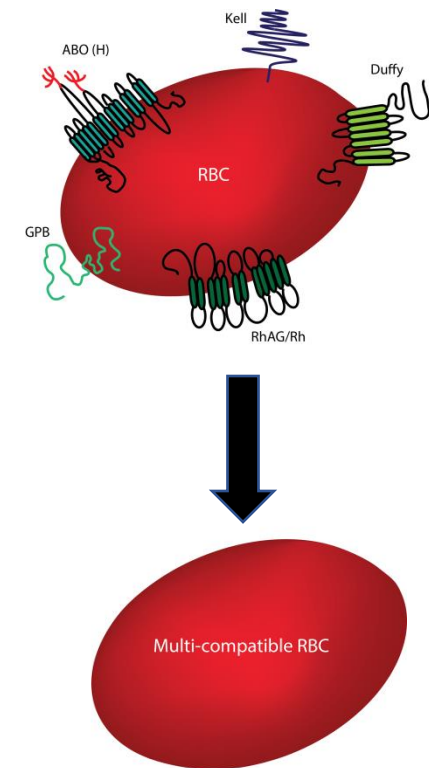
Blood Group System	Patients with Alloantibodies	Alloantibodies identified
MNS (GPB)	22	U, S, s
Rh	19	D, C, c, E, e, Hr <sup>B</sup> , hr <sup>B</sup> , Hr <sub>0</sub> , MAR, C <sup>w</sup>
Duffy	10 (+2)	Fy <sup>a</sup> , Fy <sup>b</sup> , Fy3
Kell	10	K, k, Kp <sup>a</sup>
ABO (H)	8	H (Bombay Phenotype)
Lutheran	3	Lu <sup>a</sup> , Lu <sup>b</sup>
Kidd	3	Jk <sup>b</sup>

18 patients (mainly sickle cell) have alloantibodies to more than one blood group  
 48 of 56 patients could be serviced by the removal of just 5 blood group proteins  
 Individual null phenotypes occur naturally without clinical phenotype (mild for Rh null)

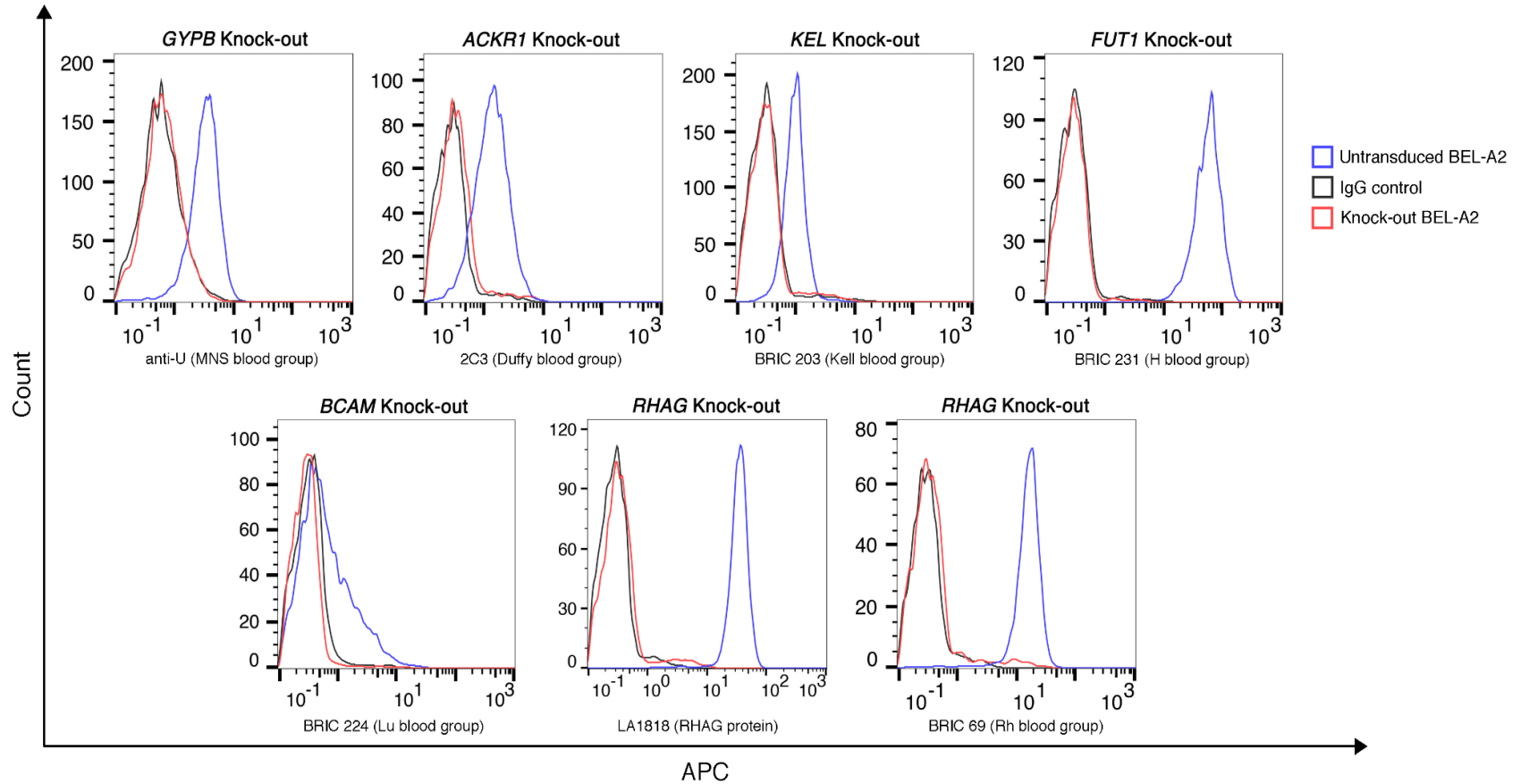
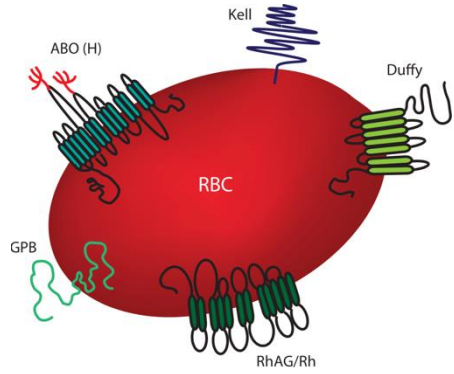
- Generate lines from individuals with relevant rare blood types
- Gene editing to customise cells with individual or combined multiple null phenotypes to broaden transfusion compatibility

(11/2014-01/2015) +  
 (05/2015-05/2016)

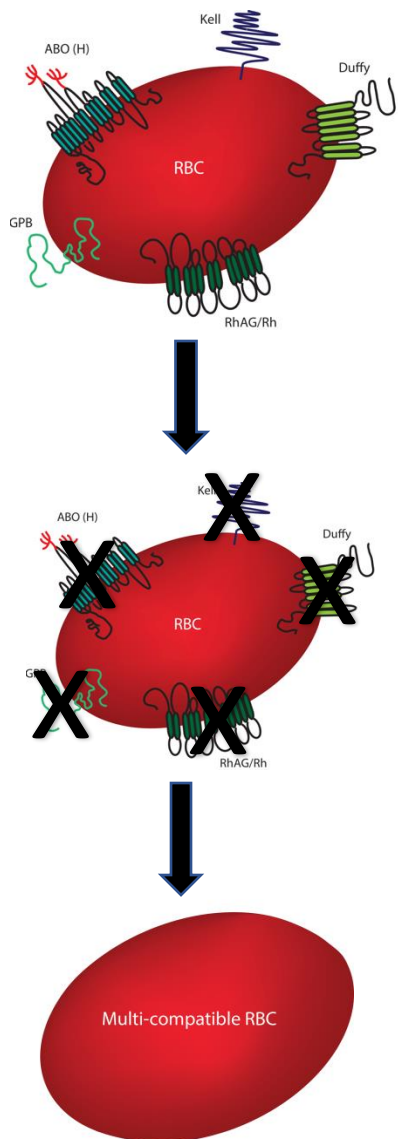
Raw data courtesy of  
 Dr. Fiona Regan NHSBT,  
 London.



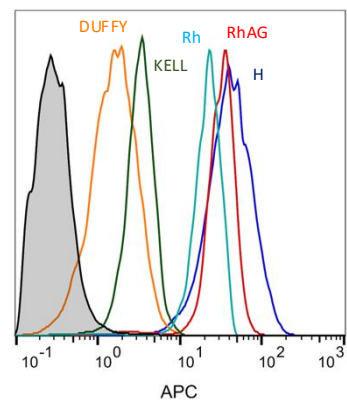
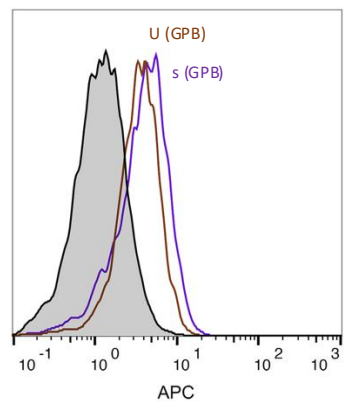
# Production of a bank of BEL-A sublines with single blood group knockouts



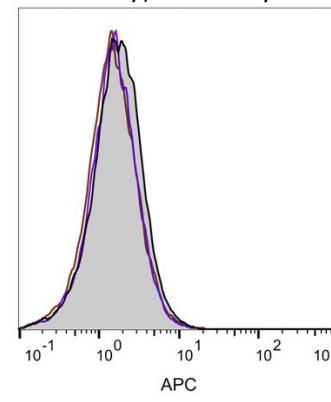
# Engineering a more compatible red blood cell



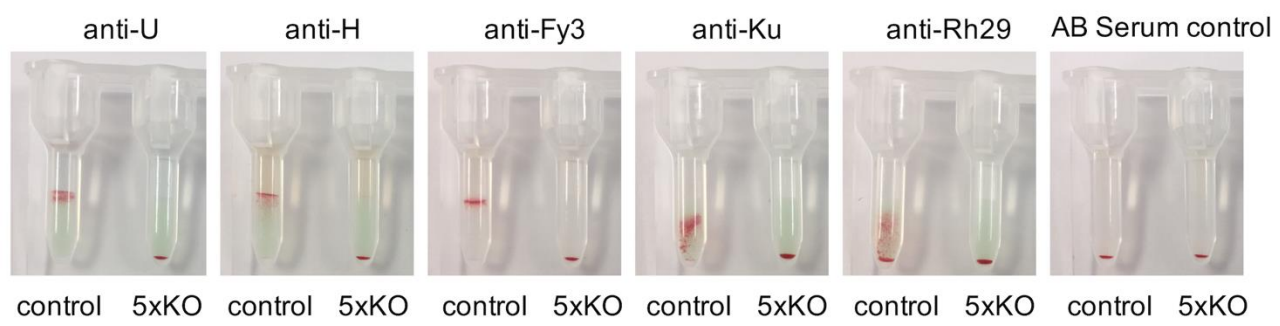
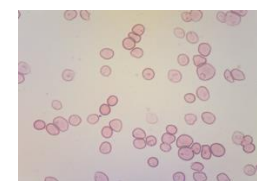
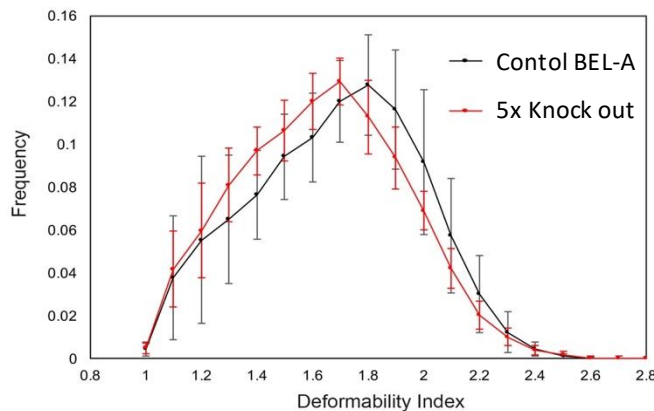
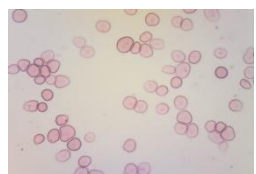
Control BEL-A Derived Reticulocytes



5x Knockout (GPB, Kell, Duffy, Rh<sub>null</sub>, Bombay) Reticulocytes

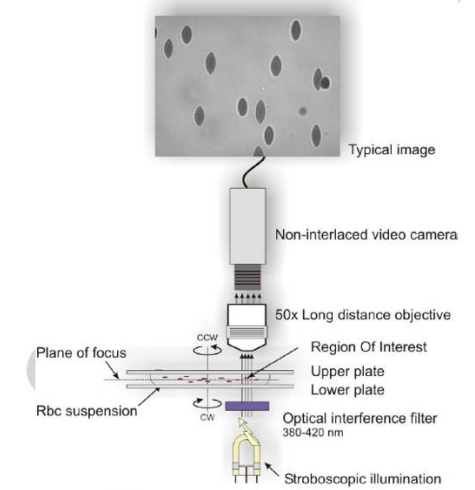


- IgG
- BRIC 203 (anti-Kell)
- LA1818 (anti-RhAG)
- 2C3 (anti-Duffy)
- BRIC 231 (anti-H)
- BRIC 69 (anti-Rh)
- anti-U
- anti-s



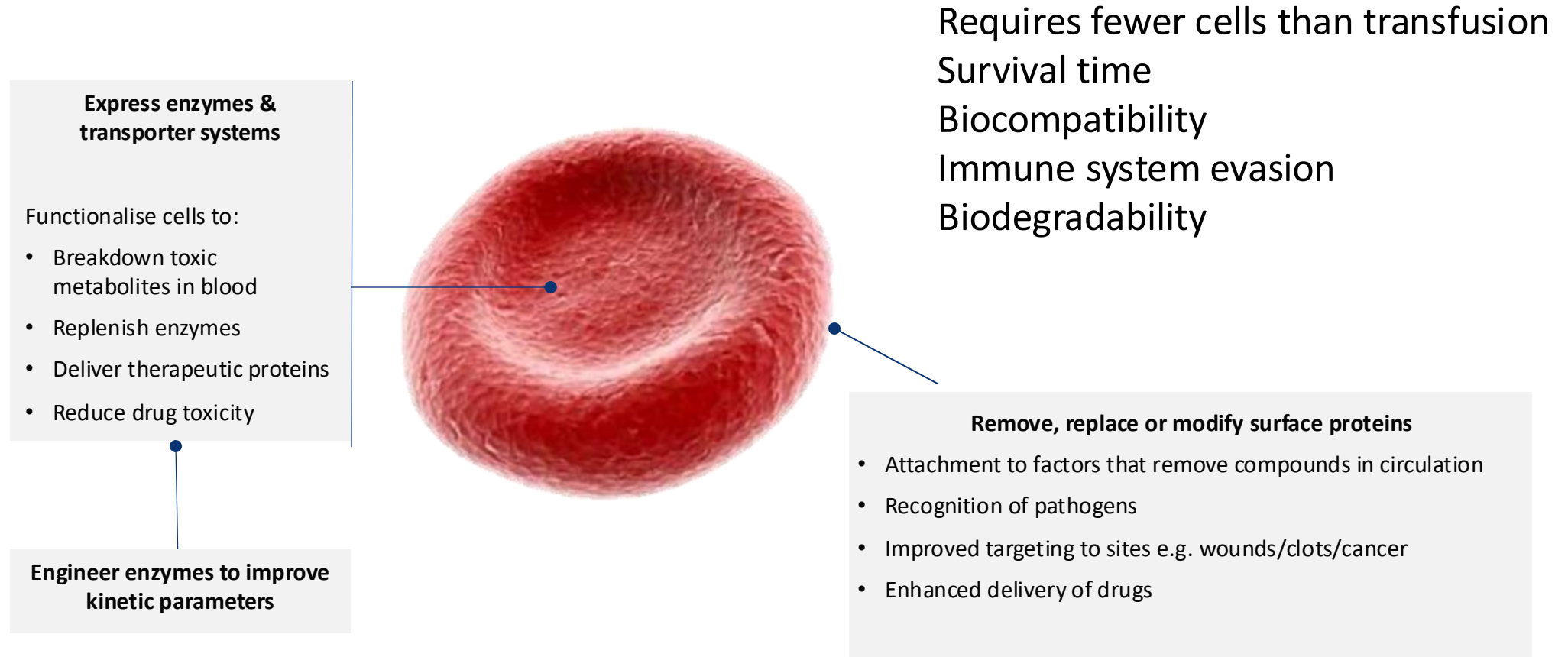
FLOW cytometry  
Comparative proteomics

Automated Rheoscope  
Cell Analyser (ARCA)



# RBCs as drug delivery and targeting agents

## Engineering RBCs to add new functionality to solve clinical problems

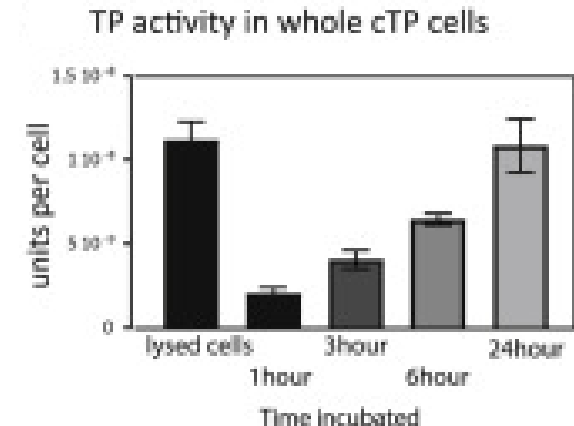
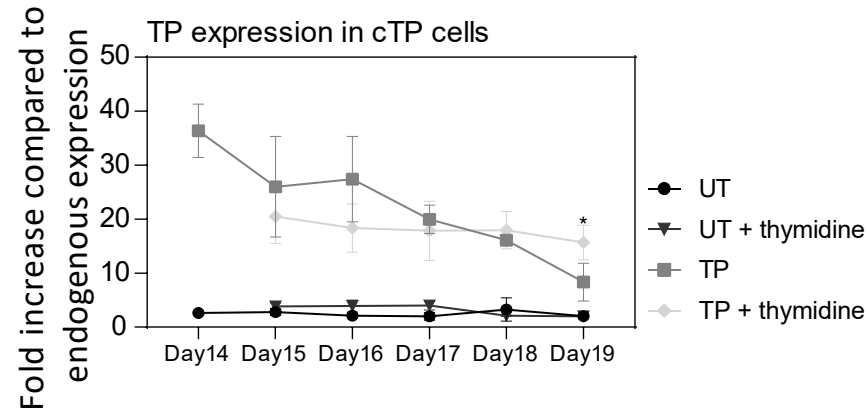
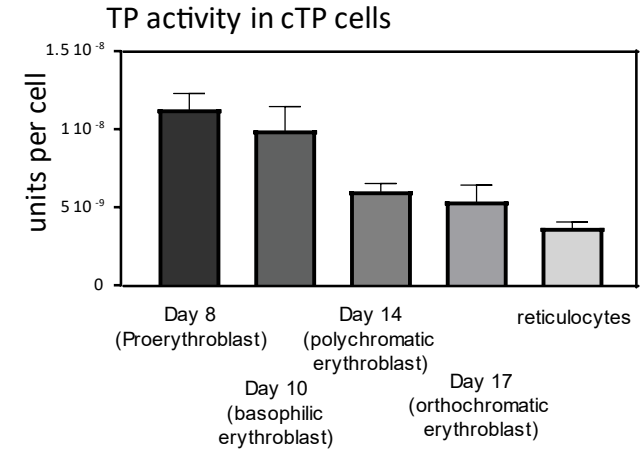
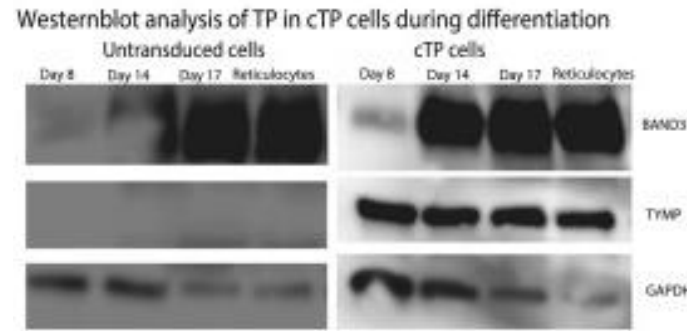


# Engineering erythroid cells to express & retain functional thymidine phosphorylase

Deficiency of thymidine phosphorylase causes Mitochondrial Gastrointestinal Encephalopathy (MNGIE)

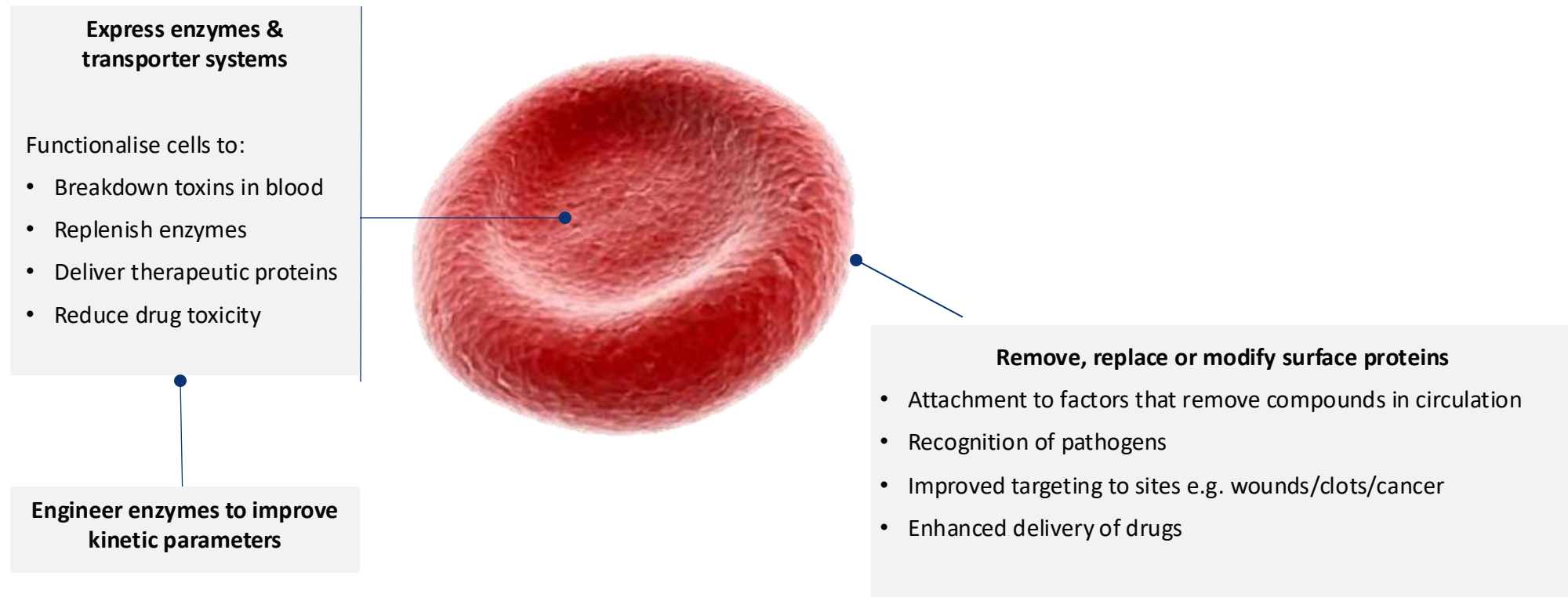
Patients have very high levels of thymidine in the body, which damages mitochondrial DNA and affects the gastrointestinal and nervous systems

**Hypothesis: expression of thymidine phosphorylase in red blood cells could break down excess thymidine**



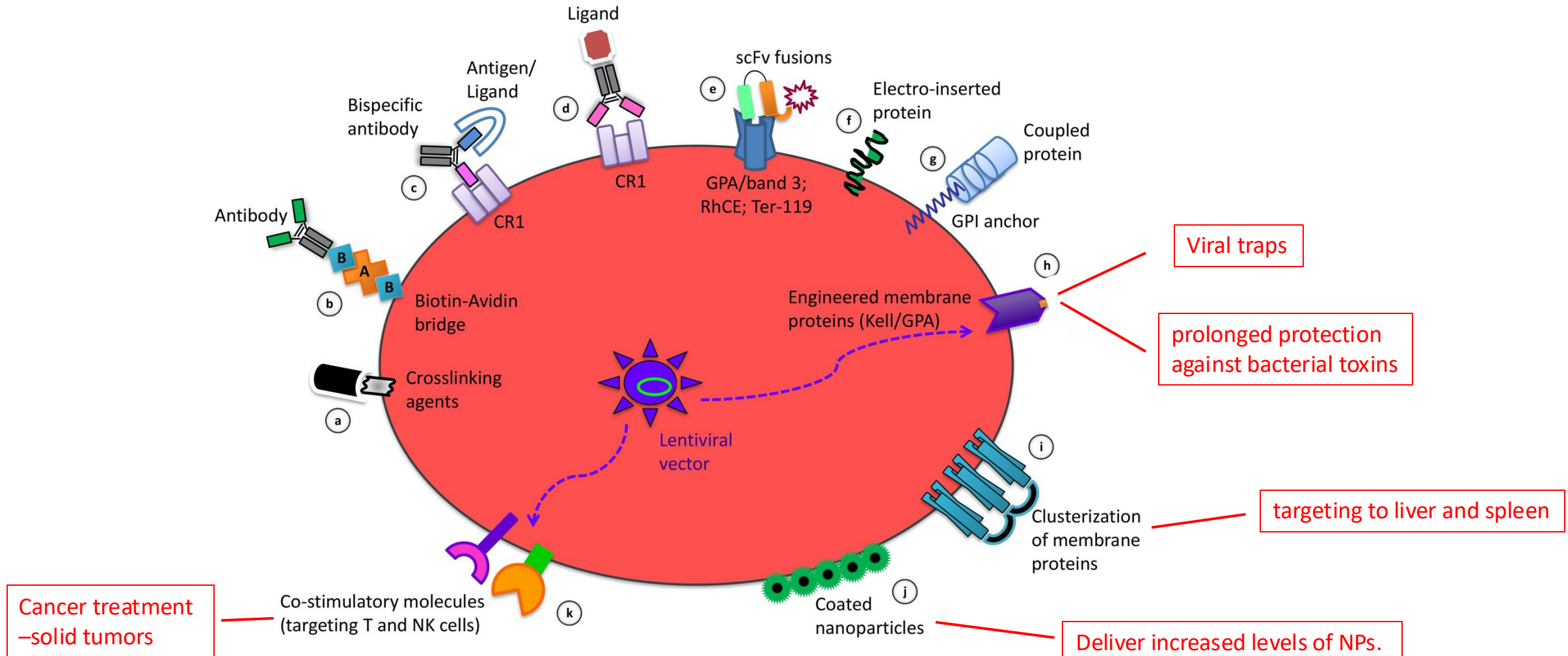
# RBCs as drug delivery and targeting agents

Engineering RBCs to add new functionality to solve clinical problems

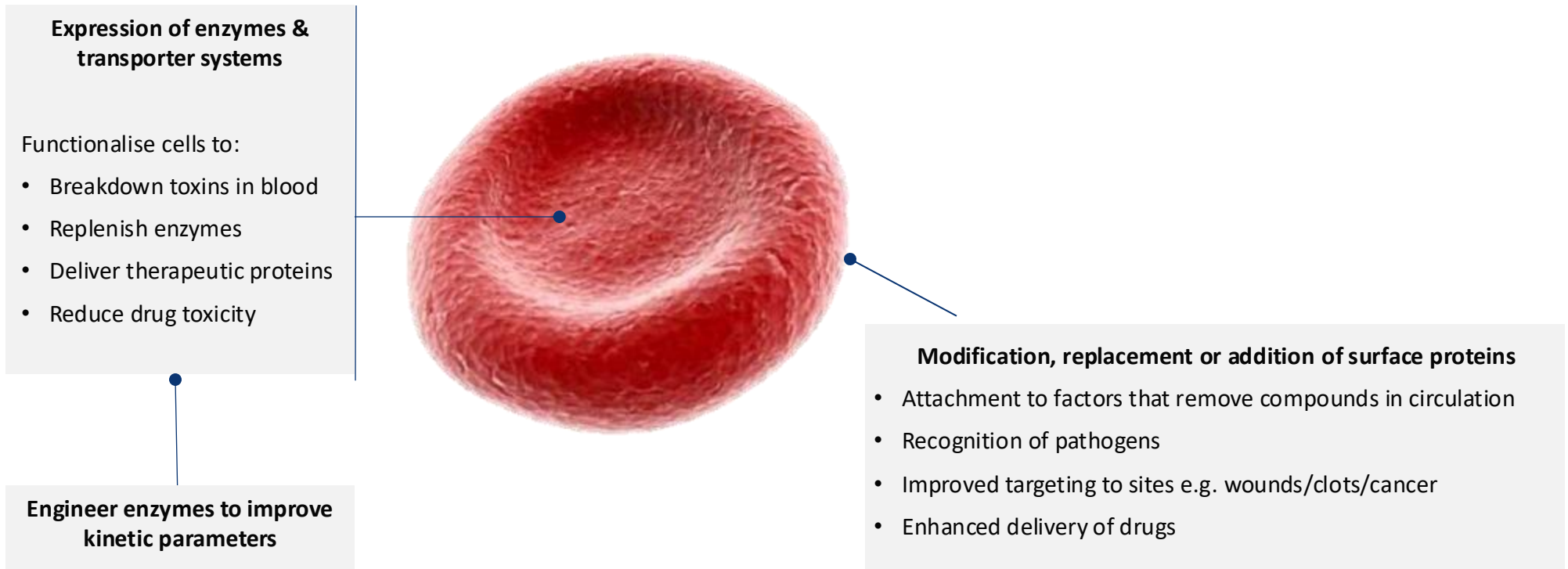


# Tagging surface of RBCs

Ectopic expression of tagged membrane protein/hybrid protein  
Tag endogenous gene using CRISPR  
Or tagging surface mature RBC



# RBCs as drug delivery and targeting agents





# Summary

- We can grow red blood cells in the laboratory, with identical features to endogenous cells
- Developed immortalised erythroid cell lines with phenotype of donor, providing a sustainable supply of banked red cells
- Genetic engineering of BEL-A to create desirable or multi-compatible red cells by gene knockout - with knock ins now possible meaning selected blood group polymorphisms can be introduced. Engineered cells can be banked
- Genetic engineering for enhanced red cell properties
- Genetic engineering to utilize red cells as drug delivery and targeting agents

# Acknowledgements

## **Frayne Lab**

Debbie Daniels  
Kontanga Trakarnsanga  
Joe Hawksworth  
Ivan Ferrer Vicens  
Daniel Ferguson

## **Toye lab**

Marjolein Meinders  
Hannah Langlands  
Tim Satchwell

## **NHSBT Filton R&D**

Sabine Kupzig  
Nicola Cogan

## **RESTORE Team**



## Nanocarriers

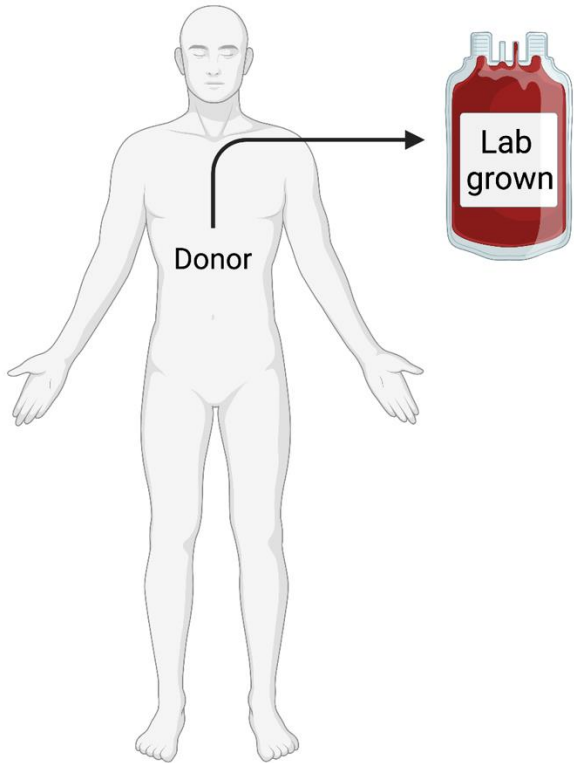
### Natural Polymers

- Chitosan
- Pullulan
- Dextran
- Hyaluronic acid
- Cycloamylose
- Glucan
- Curdlan
- Dextrin
- Gelatin
- Hydroxypropyl cellulose
- Alginate
- Heparin
- Chondroitin sulfate

### Synthetic Polymers

- Polyethylene glycol
- Polyacrylamide
- Polyglycerol
- Various polypeptides

# Donor selection for lab grown blood



Considerations:

Currently ISBT recognises ~378 blood group antigens of which 345 fall into one of ~43 Blood group systems

Type O blood is accepted as the most "Universal"

After ABO antigen the RhD antigen is most immunogenic followed by

K, E, c, Fy<sup>a</sup>, Jk<sup>a</sup> and S antigens

It has been calculated using previous blood requirements for France that 2 donor derived iPSC cell lines could theoretically cover 98.6% of patient blood needs in France\*

\*Peyrard et al Transfus Med Rev 2011